International Journal of Dentistry and Oral Health

Research Article

WILEY'S

ISSN 2471-657X

Effect of Smoking on the Expression of Human Beta-Defensin-2 in Gingival Crevicular Fluid after Non-Surgical Periodontal Therapy

Riam A. Bin Barek*, Mai S. Attia and Ossama El-Shall

Oral Medicine, Periodontology, Diagnosis and Radiology Department, Faculty of Dental Medicine, Al-Azhar University (Girls' Branch), Cairo, Egypt.

Abstract

Purpose: This study was designed to evaluate the effect of smoking on the expression of hβD-2 in the GCF after non-surgical periodontal therapy.

Methods: Ten non-smokers patients with periodontitis stage-2 grade-A and ten smoker patients with periodontitis stage-2 grade-C with age ranged between 25-40 years were selected for this study. All patients were examined with clinical periodontal parameters. Patients in both groups underwent nonsurgical periodontal therapy combined with a maintenance program (including brushing with regular toothpaste and flossing). Gingival crevicular fluid (GCF) samples were collected from all patients at baseline, one month as well as three months after periodontal therapy. Quantification of beta-defensin-2 (hβD-2) in human samples was measured using hβD-2 ELISA test.

Results: a slightly greater mean hβD-2 level at a baseline was recorded in non-smokers group (69.18±9.30) than smoker group (61.99±12.97), with no statistically significant difference (p=0.1714). At one-month, a slightly greater mean hβD-2 level was recorded in smokers group (79.90+_13.33) than non-smokers (75.89±13.41), with no statistically significant difference (p=0.0511). At three months, a slightly greater mean hβD-2 level was recorded in non-smokers group (135.77±32.83) than smoker (117.64±17.77) with no statistically significant difference (p=0.1420).

Conclusions: Non-surgical periodontal therapy resulted in relative improvement in all clinical parameters as well as an increase in hβD-2 levels. In addition, GCF levels of hβD-2 were higher after non-surgical treatment in non-smoker groups than smokers. The deficiency of hβD-2 possibly could be related to host/microbial interaction and the Smoking might modulate secretion of hβD-2, which represents a local defense dysfunction.

Keywords: Beta defensins-2; Periodontitis; Smoking

Corresponding author: Riam A. Bin Barek, M.Sc. in

Oral Medicine, Periodontology, Diagnosis and Radiology Department, Faculty of Dental Medicine, Al-Azhar University (Girls' Branch), Cairo, Egypt. E-mail: riam.2020@yahoo.com

Citation: Riam A. Bin Barek. *et al.* (2020), Effect of Smoking on the Expression of Human Beta-Defensin-2 in Gingival Crevicular Fluid after Non-Surgical Periodontal Therapy. Int J Dent & Ora Hea. 6:7

Copyright: © 2020 **Riam A. Bin Barek**. *et al*. This is an open access article distributed under the terms of the CreativeCommons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Received: September 01, 2020 Accepted: September 24, 2020 Published: October 15, 2020

1. Introduction :

Periodontitis is a longstanding disease initiated by microbial dysbiosis and characterized by activation of host-derived proteinases that lead to periodontal apparatus destruction with the subsequent further invasion of bacteria along the root surface. However, environmental factors such as smoking may modify the host's immune response to the dental biofilm so that periodontal damage becomes more progressive(1).

Smoking is one of the main and most prevalent risk factors for periodontitis. Smokers have demonstrated a decreased inflammatory response to plaque accumulation and reduced gingival bleeding(2). This altered inflammatory response has been attributed to an alteration in the gingival vasculature which includes decreased vascular density, lumen area of gingival vessels, and epithelial thickness(3,4). Furthermore, smoking produces a suppressive effect or impairment on various immune cells such as monocytes, neutrophils, lymphocytes and natural killer (NK) cells(5). In addition, smoking diminishes the phagocytic uptake of both bacteria and apoptotic cells and induces qualitative and quantitative defects in circulating NK cells which are important in host viral and anti-tumor responses(6,7). On the other hand, smoking also had a role in modulating the expression of pro-inflammatory cy-

tokines in the periodontal ligament and fibroblast cells(8)as well as in oral keratinocytes and GCF (9,10,11).

The goals of today's treatment of periodontitis are to reduce infection, resolve inflammation and create a clinical condition, which is compatible with periodontal health (12). Non-surgical periodontal therapy consists of scaling and root planing (SRP) combined with oral hygiene instructions and their efficacy directly related to the ability of the treatment to lower levels and prevalence of one or more pathogenic bacterial species. Subsequently, this results in attachment gain and pocket depth reduction due to a resolution of the inflammation(13,14,15).

Antimicrobial peptides (AMPs) are multifunctional peptides whose fundamental biological role has been proposed to be the elimination of a diverse spectrum of microorganisms(16).The most important antimicrobial peptide group in humans is defensins. Defensins have the ability to inactivate many bacteria, fungi, and some enveloped viruses. In humans, defensins can be subdivided into two families: alpha-defensins and beta-defensins(17,18).

The human beta-defensins (h β Ds) are small, cationic AMPs made primarily by epithelial cells and expressed in all human epithelia (19). The h β Ds are secreted in biological fluids, including urine, bronchial fluids, nasal secretions, saliva and GCF (20,21,22).

Among different AMPs, in the oral cavity, h β Ds-2 was found to be 10-fold more potent than h β D-1 and exhibited microbicidal activities against gram-positive and gram-negative bacteria, fungi, and some parasites (23,24,,25)Furthermore, h β D-2 mainly secreted in response to stimulation. This stimulation does not necessarily come only from bacteria since proinflammatory cytokines such as TNF- α , interferon (IFN)-gamma, IL-1 β , IL-17, and IL-22, stimulate h β D-2 secretion. Otherwise, anti-inflammatory cytokines such as IL-4, and IL-10, suppress its production(26). Besides, h β D-2 brings blood cells to the site of infection by acting as chemotactic agents(26,27). H β D-2 also was found to trigger fibroblast proliferation(28). Additionally, h β D-2 has a strong impact on the maturation of premature osteoblasts which might be effective in bone tissue regeneration (29). The present study aimed to investigate the effect of smoking on the expression of h β D-2 in the GCF after non-surgical periodontal therapy.

2. Methods:

Twenty patients (age ranged 25-40 years) were selected from those attended to the Outpatient Clinics of Oral Medicine, Periodontology, Oral Diagnosis & Radiology department, Faculty of Dental Medicine, Al-Azhar University (Girls' Branch), clinically diagnosed as having periodontitis according to the classification of periodontal diseases by(1). The criteria for inclusion in the current study were including patients free from any systemic conditions that affect the periodontium or interfere with periodontal treatment, diagnosed as having stage 2 periodontitis with CAL 5 mm or less , radiographic bone loss 15-33% and no history of periodontal tooth loss, did not receive any periodontal treatment in the past six months before the examination, and did not receive antibiotics or anti-inflammatory therapy in the six months before the examination, for female patients, no pregnancy or lactation was included.

All individuals were informed about the procedures of the study and the benefits of their participation in the study. A satisfactory written consent was obtained from all the patients denoting they're convinced about the schedule research program design. The ethical committee meeting approved the study protocol. The smoking history of the patients was evaluated using a questionnaire, after which the patients were divided into two groups based on their smoking history. If the patient smoked more than 10 cigarettes per day, then he/she was classified as a smoker; if he/she had never smoked, then he/ she was classified as a non-smoker(30).

Each patient's periodontal status was evaluated by measuring the Plaque Index (Pl)(31), Gingival Index (GI) (32)Periodontal Probing Depth (PPD), Clinic Attachment Loss (CAL); at the baseline, one-month, and at three-months intervals by using Michigan 'O' Probe With Williams graduated periodontal probe. Full mouth records were the target for recording these parameters.

Collection of samples:

Samples of GCF were collected at baseline, one-month and threemonths regarding the periodontal therapy. The samples were pooled from four periodontal sites with attachment loss of 4-5mm(in the four different quadrants). The sampling area was isolated with cotton rolls and carefully cleaned supragingivally with sterile cotton pellets. A sterile absorbent paper point was inserted into the gingival crevice or pocket until resistance was felt. The paper point was held in place for the 30s. The samples were immediately placed in Eppendorf tubes, transported to the laboratory and stored at -80°C. The collected samples analyzed using the enzyme-linked immunosorbent assay (ELISA) technique of human beta defensin-2 kit.

Non-Surgical Periodontal Therapy:

All patients in both groups were treated with nonsurgical periodontal therapy, which included the following: Supragingival and subgingival scaling and root debridement were performed with an ultrasonic device using iPiezo engine (NSK Varios 970, Japan), Chlorhexidine mouthwash was prescribed twice daily for one week post periodontal therapy and Oral hygiene instructions included brushing teeth with soft dental brush three times daily and using dental floss once a day.

Quantification of Human beta defensin-2 using ELISA technique:

Quantification of h β D-2 in human samples was measured using Bioneovan Inova h β D-2 ELISA kit. The kit is suitable for testing a variety of sample types in-vitro and Purchased from Bioneovan Inova Co. Beijing, China. The kit assayed h β D-2 level in the sample, using a Purified h β D-2 antibody to coat microtiter plate wells, made a solid-phase antibody, then added h β D-2 to wells, Combined HBD2 antibody which With HRP labeled, become antibody-antigen - enzyme-antibody complex. After washing Completely, Added TMB substrate solution, TMB substrate becomes blue color At HRP enzyme-catalyzed, reaction is terminated by the addition of a sulphuric acid solution and the color change is measured spectrophotometrically at a wavelength of 450 nm. The concentration of h β D-2 in the samples is then determined by comparing the O.D. of the samples to the standard curve.

Statistical analysis:

Values were presented as mean and standard deviation (SD) values. Data were explored for normality using Kolmogorov-Smirnov test of normality. The results of Kolmogorov-Smirnov test indicated that most of the data were normally distributed (parametric data), so oneway analysis of variance ANOVA test was used to compare between different intervals within the same group, followed by Tukey's post hoc test when the difference was found to be significant. Unpaired

t-test was used to compare both groups (non-smokers, smokers). The significance level was set at p \leq 0.05. Statistical analysis was performed with SPSS 16.0 (Statistical Package for Scientific Studies, SPSS, Inc., Chicago, IL, USA) for Windows.

3. Results:

Table (1) showed the changes in the scores and measurements of PI, GI, PPD and CLA at baseline, 1-month and 3-months after non-surgical periodontal therapy in the periodontitis of smoker and non-smoker patients.

 $H\beta$ Ds-2 ELISA analysis showed that in both groups, $h\beta$ D-2 level increased after non-surgical therapy, to reach the highest mean value after 3 months (Table-2, Figure-1). One-way analysis of variance revealed that there is a statistically significant increase by time in both groups (p<0.0001). Tukey's post hoc test revealed no significant differ-

ence between mean values recorded at baseline (69.18 ± 9.30), after one month (75.89 ± 13.41) and after three months (135.77 ± 32.83) in the non-smokers group. However, in the smokers group, there was a significant difference between baseline (61.99 ± 12.97), after one-month (79.90 ± 13.33) and after three month observation times (117.64 ± 17.77).

At a baseline, a slightly greater mean h β D-2 level was recorded in non-smokers group (69.18±9.30) than smoker group (61.99±12.97), with no statistically significant difference (p=0.1714). At one-month, a slightly greater mean h β D-2 level was recorded in smokers group (79.90+_13.33) than non-smokers (75.89±13.41), with no statistically significant difference (p=0.0511). At three months, a slightly greater mean h β D-2 level was recorded in non-smokers group (135.77±32.83) than smoker (117.64±17.77) with no statistically significant difference (p=0.1420).

Table 1. Clinical parameters at baseline, 1-month and at 3-month evaluation

Parameter	Non-smoking group	Smoking group	P value
PI			
Baseline	1.63± 0.53	1.68± 0.55	P<0.05
1 month	0.37± 0.19	0.28± 0.09	
3 month	0.01+0.006	0.02± 0.003	
P value	<0.0001	<0.0001	
GI			
Baseline	1.53± 0.57	1.01± 0.65	P<0.05
1 month	0.36± 0.27	0.27± 0.15	
3 month	0.01± 0.004	0.03± 0.01	
P value	<0.0001	<0.0001	
PPD (mm)			
Baseline	3.17± 0.50	3.33± 0.48	P<0.05
1 month	2.55± 0.52	3.01± 0.42	
3 month	1.84± 0.54	2.53± 0.51	
P value	<0.0001	<0.0001	
CAL (mm)			
Baseline	3.77± 0.70	3.74± 0.57	P<0.05
1 month	3.11± 0.78	3.42± 0.52	
3 month	2.42± 0.75	2.94± 0.57	
P value	<0.0001	<0.0001	

Table 2. hbD-2 levels in both groups and significance of the difference between groups using unpaired t test

Groups	Baseline		1 month		3 months	
	Non-smokers	Smokers	Non-smokers	Smokers	Non-smokers	Smokers
Mean	69.18±9.30	61.99±12.97	75.89±13.41	79.90±13.33	135.77±32.83	117.64±17.77
	1.4246		0.6707		1.5358	
P value	0.1714 ^{ns}		0.0511 ^{ns}		0.1420 ^{ns}	

Ns=non-significant at p<0.05



Figure (1): Column chart showing mean hβD-2 levels in both groups

4.Discussion:

The complex effects of smoking on periodontal and oral diseases, and the mechanisms that mediate these diseases, are still considered important. However, establishing a link between cigarette smoking and abnormal levels of antimicrobial peptides will provide new insight into the epidemiology of the less favorable response following non-surgical periodontal therapy.

Human beta defensin-2 considered one of the important AMPs in epithelial innate immunity, and their differential expression is associated with periodontal health and diseases. The h β D-2 has a significant role as chemotactic, trigger fibroblast proliferation and has a strong impact on the maturation of premature osteoblasts, which might be effective in bone tissue regeneration. As such, the understanding of their role will undoubtedly unfold their clinical application in periodontal diseases.

According to(33) the expression level of h β D-2 and h β D-3 in GCF among the smoking group was significantly lower than that in the non-smoking group. Also, The mRNA expression level of h β D-2 and h β D-3 in the smoking group was weakened compared with that in the non-smoking group indicating that smoking may have a negative effect on the immune defense system of the periodontal host, however, this study is in agreement with another study by(34) demonstrating that the whole cigarette smoke (WCS) exposure remarkably attenuated h β D-3 expression levels, suggesting a link between cigarette smoke and abnormal levels of antimicrobial peptides.

The current study demonstrated that after non-surgical periodontal therapy in both groups, h β D-2 level increased after non-surgical periodontal therapy, to reach the highest mean value after 3 months. However, at baseline, a slightly greater mean h β D-2 levelwas recorded in non-smokers group, with no statistically significant difference. At one-month, a slightly greater mean h β D-2 level was recorded in smokers group, with no statistically significant difference. At three months greater mean h β D-2 level was recorded in non-smokers group, with no statistically significant difference. At three months greater mean h β D-2 level was recorded in non-smokers group, with no statistically significant difference.

At baseline, the low level of h β D-2 in smoker's group could be explained by the fact that periodontopathogenic microorganisms mainly P.gingivalis, which had a specific role in β -defensins degradation was

found in smokers more than in non-smokers(35,36). Moreover, bacteria with resistance to β -defensins, such as T. denticola and P.gingivalis, survive and colonize on epithelial surfaces, and eventually, invade gingival tissues(37). With the bacterial invasion, β -defensins stimulate the secretion of chemokines, such as IL-8 and MCP-1, from dendritic cells, and, also, act as chemoattractants, which bring phagocytes and lymphocytes to the site of infection. Correspondingly, the activated immune response limits innate response and, hence, secretion of β -defensins (38). Our findings were in agreement with previous studies shown that smoking down regulates h β D-2 expression33,35,39. On the other hand, the proteolytic enzymes, which are produced by periodontal pathogens and the host in different ways such as (trypsin-like proteases and gingipains of P.gingivalis) potentially degrade and inactivate h β D-2 in-vitro conditions 40,41-43.

The improvement noticed in terms of clinical parameters after one and three months of non-surgical periodontal therapy when compared to baseline could be explained by the positive effect of non-surgical periodontal therapy suggesting the relationship between host/bacterial factors of periodontitis and h β D-2 levels in GCF.

5.Conclusion:

The GCF levels of human h β D-2 among stage 2 periodontitis patients could be changed depending on some factors such as smoking. Smoking might also affect the different clinical parameters including; Pl, Gl, PD and CAL. Moreover, the non-surgical periodontal therapy may lead to increased levels of h β D-2 in GCF among both smokers and non-smokers. However, The discrepancies of h β D-2 slightly greater in the smoker group after therapy, which could represent, a local defense dysfunction.

References:

1. Tonetti MS, Greenwell, and Kornman KS. Staging and grading of periodontitis: Framework and proposal of a new classification and case definition. Journal of Clin Periodontol. 2018; 45 Suppl 20: S149-S161.

2. Leite FR, Nascimento GG, Scheutz F, and Lopez R. Effect of smoking on periodontitis: a systematic review and meta-regression. American journal of preventive medicine. 2018; 54 (6): 831-841.

3.BERGSTRÖM J, PERSSON L, and PREBER H. Influence of cigarette

smoking on vascular reaction during experimental gingivitis. European Journal of Oral Sciences. 1988; 96: 34-39.

4. Villar CC and de Lima AF. Smoking influence on the thickness of marginal gingival epithelium. Pesqui Odontol Bras. 2003; 17 (1): 41-45.

5. Mehta H, Nazzal K, and Sadikot RT. Cigarette smoking and innate immunity. Inflammation Research. 2008; 57: 497-503.

6. Ginns L C, Ryu JH, Rogol PR, Sprince NL, Oliver LC, and Larsson CJ. Natural killer cell activity in cigarette smokers and asbestos workers. American Review of Respiratory Disease.1985; 131: 831-834.

7. Lu LM, Zavitz CC, Chen B, Kianpour S, Wan Y, and Stampfli MR. Cigarette smoke impairs NK cell- dependent tumor immune surveillance. Journal Immunology. 2007; 178 (2): 936-943

8. Alpar B, Leyhausen G, Sapotnick A, Gunay H, and Geurtsen W. Nicotine-induced alterations in human primary periodontal ligament and gingiva fibroblast cultures. Clinical Oral Invest. 1998; 2 (1): 40-46.

9. Petropoulos G, McKay IJ, and Hughes FJ. The association between neutrophil numbers and interleukin-1 α concentrations in gingival crevicular fluid of smokers and non-smokers with periodontal disease. Journal of Clinical Periodontology. 2004; 31: 390-395.

10. Tymkiw, KD, Thunell DH, Johnson GK, *et al.* Influence of smoking on gingival crevicular fluid cytokines in severe chronic periodontitis. Journal of Clinical Periodontology 2011; 38: 219-228.

11. Johnson GK, Guthmiller JM, Joly S, Organ CC, and Dawson DV. Interleukin-1 and interleukin-8 in nicotine- and lipopolysaccharide-exposed gingival keratinocyte cultures. Journal of Periodontal Research. 2010; 45 (4): 583-588.

12. Lang NP and Tonetti MS. Periodontal risk assessment (PRA) for patients in supportive periodontal therapy (SPT). Oral Health Prev Dent. 2003; 1(1): 7-16.

13. Stelzel M, Florèsde Jacoby L. Topical metronidazole application as an adjunct to scaling and root planing. Journal of Clinical Periodontology. 2000;27(6):447-452.

14. Listgarten M, Lindhe J, and Hellden L. Effect of tetracycline and/ or scaling on human periodontal disease: Clinical, microbiological, and histofogical observations. Journal of Clinical Periodontology. 1978; 5(4): 246-271.

15. Badersten A, Nilvéus R, and Egelberg J. Effect of nonsurgical periodontal therapy: I. Moderately advanced periodontitis. Journal of Clinical Periodontology. 1981;8(1):57-72.

16. Diamond G, Beckloff N, Weinberg A, and Kisich KO. The roles of antimicrobial peptides in innate host defense. Current Pharmaceutical Design. 2009;15(21):2377-2392.

17. Scott MG and Hancock RE. Cationic antimicrobial peptides and their multifunctional role in the immune system. Critical Reviews[™] in Immunology. 2000; 20 (5).

18. Froy O. Microreview: Regulation of mammalian defensin expression by Toll like receptor dependent and independent signalling pathways. Cellular microbiology. 2005; 7 (10): 1387-1397.

19. Dale BA. Periodontal epithelium: a newly recognized role in health and disease. Periodontology 2000. 2002;30 (1): 70-78.

20. Valore EV, Park CH, Quayle AJ, Wiles KR, McCray PB, and Ganz T. Human beta-defensin-1: an antimicrobial peptide of urogenital tissues. The Journal of Clinical Investigation. 1998; 101(8): 1633-1642.

21. Sahasrabudhe K, Kimball J, Morton T, Weinberg A, Dale B. Expression of the antimicrobial peptide, human β -defensin 1, in duct cells of minor salivary glands and detection in saliva. Journal of Dental Research. 2000; 79 (9): 1669-1674.

22. Diamond DL, Kimball JR, Krisanaprakornkit S, Ganz T, and Dale BA. Detection of β -defensins secreted by human oral epithelial cells. Journal of immunological methods. 2001;256(1-2):65-76.

23. Yang D, Liu Z-h, Tewary P, Chen Q, De la Rosa G, and Oppenheim JJ. Defensin participation in innate and adaptive immunity. Current pharmaceutical design. 2007; 13 (30): 3131-3139.

24. Singh PK, Jia HP, and Wiles K, *et al.* Production of β -defensins by human airway epithelia. Proceedings of the National Academy of Sciences. 1998;95(25):14961-14966.

25. García J-RC, Krause A, Schulz S, *et al*. Human β -defensin 4: a novel inducible peptide with a specific salt-sensitive spectrum of antimicrobial activity. The FASEB Journal. 2001;15(10):1819-1821.

26. Kanda N, Kamata M, Tada Y, Ishikawa T, Sato S, and Watanabe S. Human β defensin 2 enhances IFN γ and IL10 production and suppresses IL17 production in T cells. Journal of leukocyte biology. 2011; 89 (6): 935-944.

27. Pazgier M, Hoover D, Yang D, Lu W, and Lubkowski J. Human β -defensins. Cellular and Molecular Life Sciences CMLS. 2006; 63 (11): 1294-1313.

28. Nishimura M, Abiko Y, Kurashige Y, *et al*. Effect of defensin peptides on eukaryotic cells: primary epithelial cells, fibroblasts and squamous cell carcinoma cell lines. Journal of dermatological science. 2004;36(2):87-95.

29. Kraus D, Deschner J, Jäger A, *et al*. Human β -defensins differently affect proliferation, differentiation, and mineralization of osteoblastlike MG63 cells. Journal of cellular physiology. 2012; 227 (3): 994-1003.

30. Ertugrul AS, Sahin H, Dikilitas A, Alpaslan NZ, Bozoglan A, Tekin Y. Gingival crevicular fluid levels of human beta-defensin-2 and cathelicidin in smoker and non-smoker patients: a cross-sectional study. Journal of Periodontal Research. 2014;49(3):282-289.

31. Silness J and Löe H. Periodontal disease in pregnancy II. Correlation between oral hygiene and periodontal condition. Acta odontologica scandinavica. 1964; 22 (1): 121-135.

32.Löe H and Silness J. Periodontal disease in pregnancy I. Prevalence

and severity. Acta odontologica scandinavica. 1963; 21 (6): 533-551.

33. Fan Y, Ye D, Zhou X, Yu F, Li W. The impact of smoking on human beta defensin 2, 3 in gingival crevicular fluid and gingival tissue of patients with chronic periodontitis. Shanghai Journal of Stomatology. 2015;24(6):735-738.

34. Wang W-m, Ye P, Qian Y-j, *et al*. Effects of whole cigarette smoke on human beta defensins expression and secretion by oral mucosal epithelial cells. Tobacco induced diseases. 2015; 13(1): 3.

35. Mahanonda R, SaArd Iam N, Eksomtramate M, *et al.* Cigarette smoke extract modulates human β -defensin-2 and interleukin-8 expression in human gingival epithelial cells. Journal of periodontal research. 2009;44(4):557-564.

36. Zambon J, Grossi S, Machtei E, Ho A, Dunford R, and Genco R. Cigarette smoking increases the risk for subgingival infection with periodontal pathogens. Journal of Periodontology. 1996; 67: 1050-1054.

37. Brissette C, Simonson L, and Lukehart S. Resistance to human β defensins is common among oral treponemes. Oral microbiology and immunology. 2004; 19(6): 403-407.

38. Yin L, Chino T, Horst OV, *et al*. Differential and coordinated expression of defensins and cytokines by gingival epithelial cells and dendrit-

ic cells in response to oral bacteria. BMC Immunology. 2010; 11(1): 37.

39. Wolgin M, Liodakis S, Pries AR, Zakrzewicz A, and Kielbassa AM. HBD-1 and hBD-2 expression in HaCaT keratinocytes stimulated with nicotine. Archives of Oral Biology. 2012; 57(6): 814-819.

40. Taggart CC, Greene CM, Smith SG, *et al.* Inactivation of human β -defensins 2 and 3 by elastolytic cathepsins. The Journal of Immunology. 2003; 171(2): 931-937.

41. Kuula H, Salo T, Pirilä E, et al. Human β -defensin-1 and-2 and matrix metalloproteinase-25 and-26 expression in chronic and aggressive periodontitis and in peri-implantitis. Archives of oral biology. 2008;53(2):175-186.

42. Brancatisano FL, Maisetta G, Barsotti F, *et al*. Reduced human beta defensin 3 in individuals with periodontal disease. Journal of Dent. Research. 2011; 90(2): 241-245.

43. Maisetta G, Brancatisano FL, Esin S, Campa M, and Batoni G. Gingipains produced by Porphyromonas gingivalis ATCC49417 degrade human- β -defensin 3 and affect peptide's antibacterial activity in vitro. Peptides. 2011; 32(5): 1073-1077.